

# DNA SEQUENCE DATA

-From template DNA to  
Sequence Alignment...

## **Case Study:** **Western Diamondback Rattlesnake (*Crotalus atrox*)**

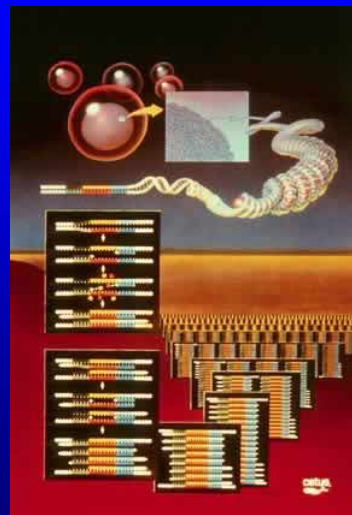


## Protocol

1. Collect tissue samples from *C. atrox* individuals and extract tDNA
2. Amplify specific gene using PCR (Polymerase Chain Reaction)
3. Sequence PCR products
4. Align our sequence with published sequences
5. Analyze with phylogenetic software

## PCR – Purpose

- Need multiple copies of the gene in order to sequence it
- Primer extension reaction for amplification of specific nucleic acids in vitro



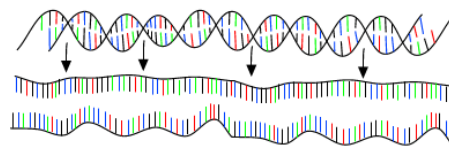
# PCR – Reaction Composition

- tDNA
- Sequence specific primers
- dNTP's
- Taq polymerase
- Buffer
- Thermocycler



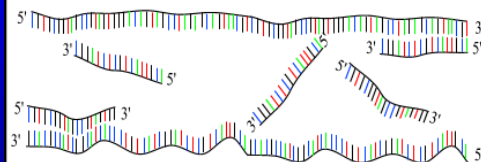
## PCR : Polymerase Chain Reaction

30 - 40 cycles of 3 steps :



Step 1 : denaturation

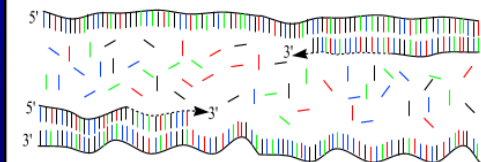
1 minut 94 °C



Step 2 : annealing

45 seconds 54 °C

forward and reverse primers !!!

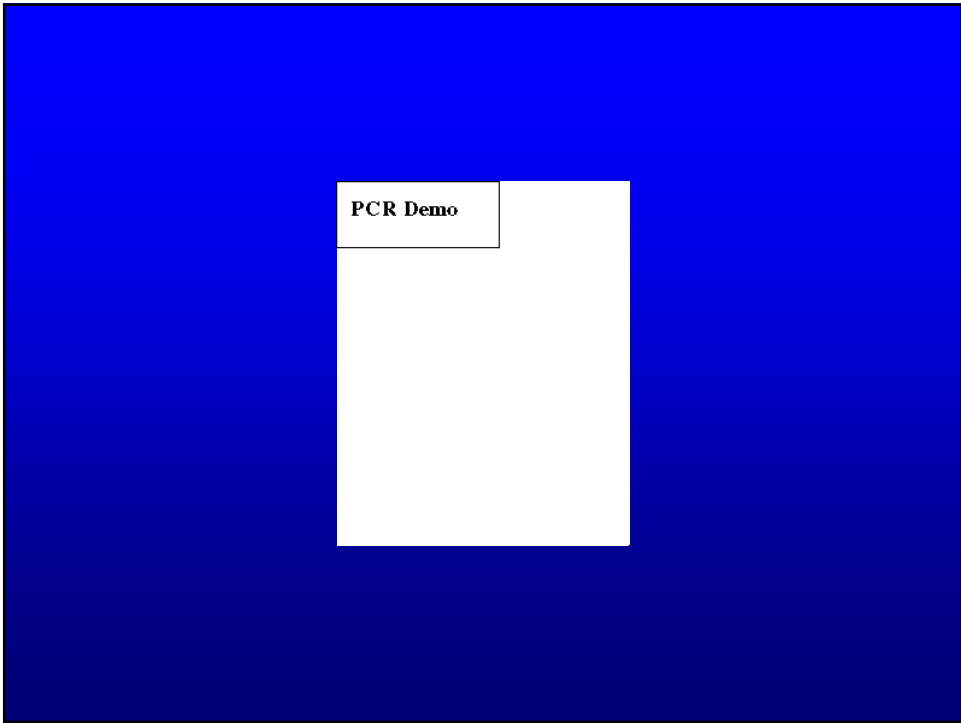
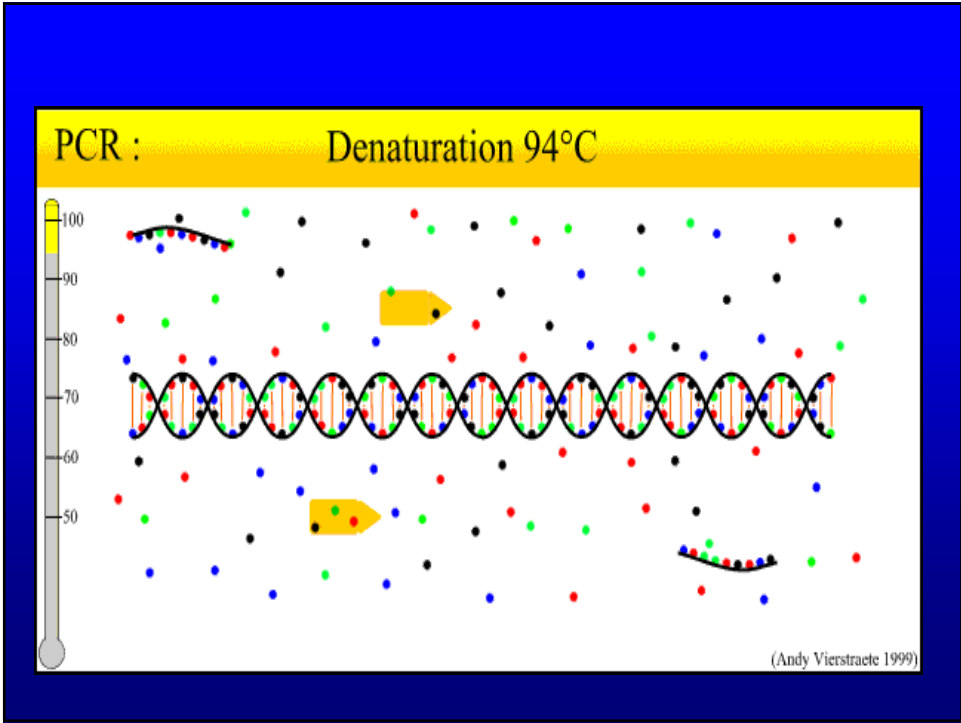


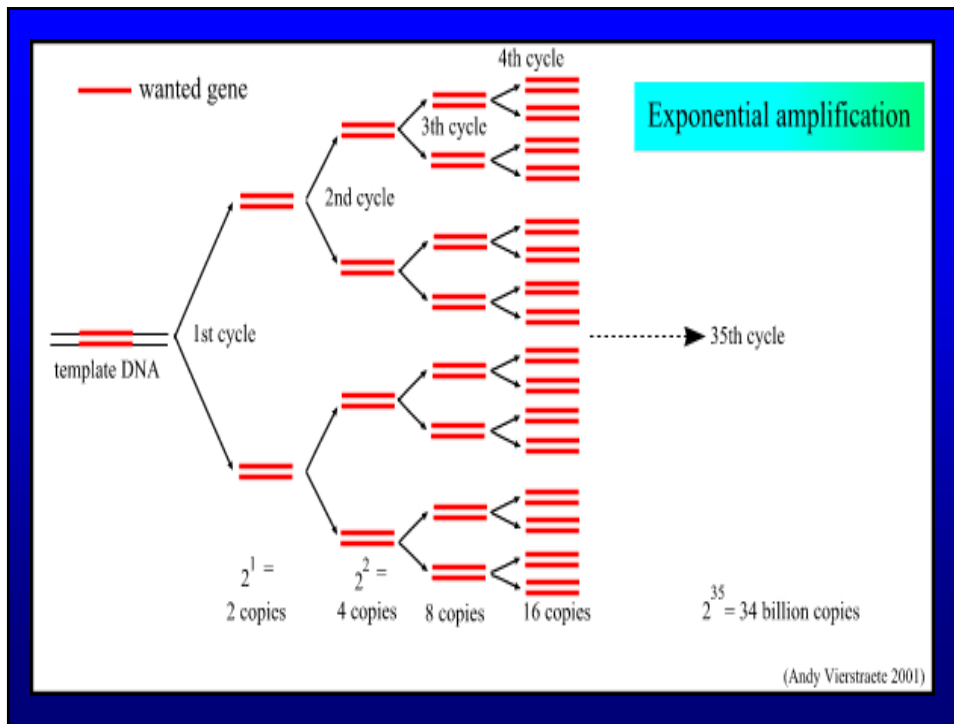
Step 3 : extension

2 minutes 72 °C

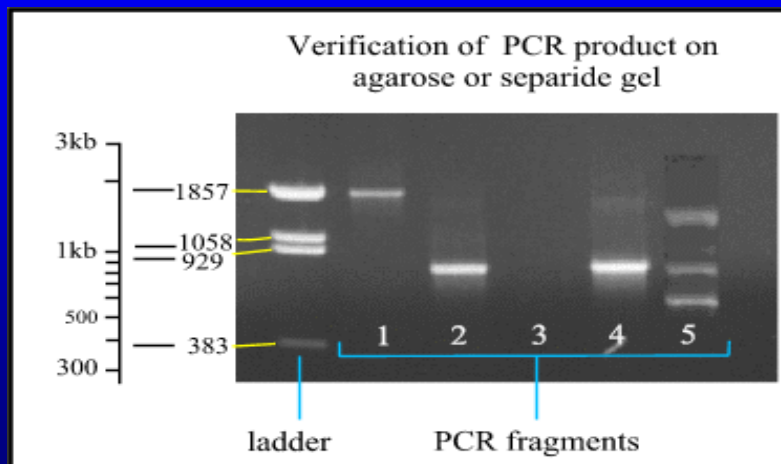
only dNTP's

(Andy Vierstraete 1999)

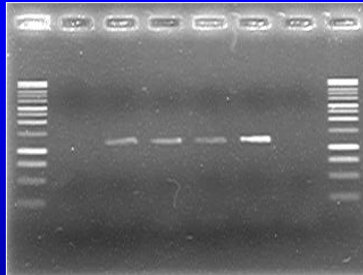




## PCR – How do we know it worked?



# DNA Sequencing



How to get  
from this...

To this!

```
TATCGCATAATACAGATCCTCCCAACAAAACCGACCTATTCGTTGCATTGATCAT  
TGTAGCCCTCTGAGGGCAATTGTAGCCAAATCTGACATGCCTAGAACAGACAGACTAA  
ATCCCTAATCGCCTACTCCTCCATCAGCCAGATAGGGCTAGTAGAGCCGAATTTATAT  
CCAAACCCCATGAGGCCTATCCGGAGCCATAGCTCTAATAATCGCACACCGATTACCTC  
CTCAGCACTCTTTGCTGCTAGCTAACACACCCTATGAACGAACACACACCCGAGTCCTAAT  
TCTTACAGGAGGATTCCACAATATCCTACCCATAGCTACAACTGATGACTAGTAACAAA  
CCTCATAAACATCGCCATCCCCCTCCATAAACTTCACCGGAGAGCTCCTAATTATATC  
CGCCCTATTTAACTGATGCCAACAAACATCATCATACTAGGAATATCAATACTTATCAC  
CGCCTCTACTCCCTACATATATTTCTGTCAACACAAATAGGGCCAACTCTACTAAACAA  
CCAAAGAGAACCACACACTCCGGAGAACACTACTAATAACCCCTCCACTTGCCTCCCT  
ACTTATGATCTCCCTCAACCAAGATTACTGATCAGGAGTGTGCTAATTTAAACAAAT  
ATCAAGCTGTGACCTTGAATAAGATTAACTCGACACCGGAGAGTCCAGAGAAGCTG  
CTAACTCTTCAATCTGGCGAA-CACACCAGCCCTCTCTTCTATCAAAGGAGAATAGTTA-  
CCCGCTGGTCTTAGGCACCACTCTTGGTCAAAT
```

## Automated DTCS

(Dye Terminator Cycle Sequencing)

- Typically provides accurate reads of 600-800 b.p.
- For long fragments, two or more sequencing reactions are run
- Up to 96 run at once in a plate
- Reaction is similar to a PCR reaction, but there is no logarithmic replication, so technically a primer extension reaction



## Components

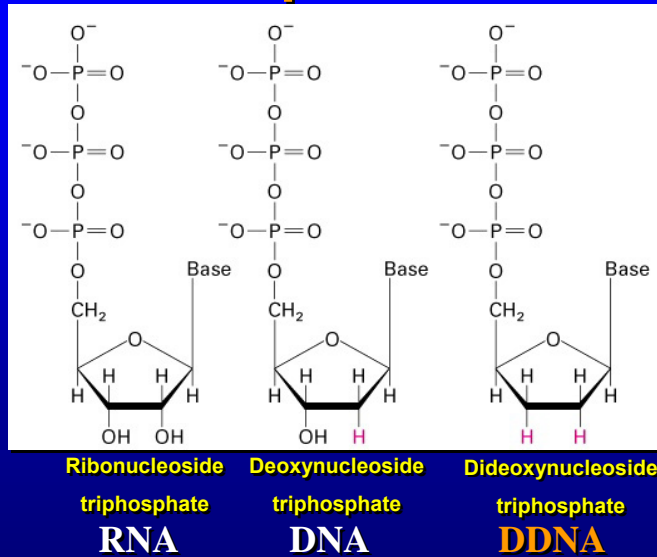
- Purified PCR product (template)
- Primer (1 per sequencing reaction)

## Components

- Thermostable DNA polymerase
- Buffer,  $MgCl_2$
- Deoxynucleoside triphosphates (dNTPs)
- Dideoxynucleoside triphosphates (ddNTPs)
  - Each with a different fluorescent label
  - Much smaller molar concentration than dNTPs



## Components



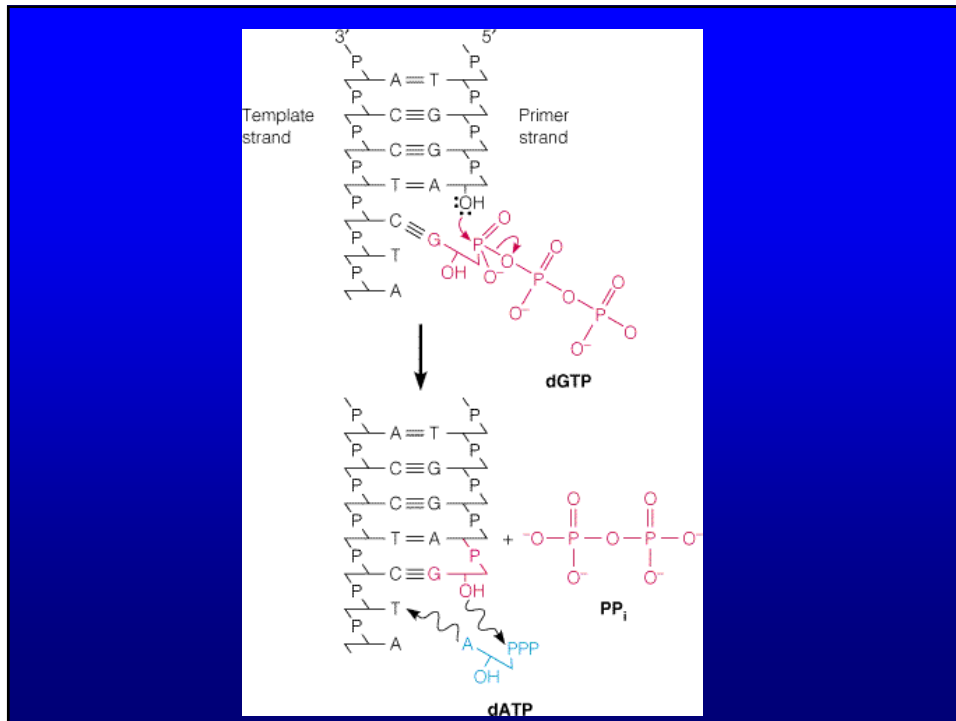
## Reaction

- **Similar to a PCR reaction:**
  - Denature at  $\sim 96^{\circ}\text{C}$
  - Anneal primer at  $\sim 50^{\circ}\text{C}$
  - Extend primer at  $\sim 60^{\circ}\text{C}$
- **Primer extension occurs normally as long as dNTPs are incorporated**
- **When a ddNTP is incorporated, extension stops**



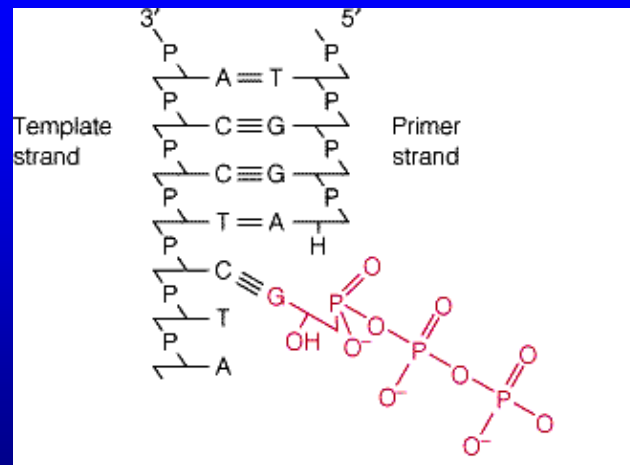
# Reaction

- **Extension occurs via nucleophilic attack**
  - 3'-hydroxyl group at the 3' end of the growing strand
  - attacks the 5'- $\alpha$ -phosphate of the incoming dNTP,
  - releasing pyrophosphate ( $PP_i$ ).
  - $(dNMP)_n + dNTP \rightarrow (dNMP)_{n+1} + PP_i$
  - Catalyzed by DNA polymerase
  - Synthesis occurs 3'  $\rightarrow$  5'



# Reaction

- ddNTPs lack a 3'-OH group
- Once a ddNTP is incorporated, nucleophilic attack cannot occur, so primer extension is terminated



[http://www.lsic.ucla.edu/l3/tutorials/gene\\_cloning.html](http://www.lsic.ucla.edu/l3/tutorials/gene_cloning.html)

## Reaction

- Produces a mixture of single-stranded DNA products of varying lengths
  - Each ends with a dye-labelled ddNTP
  - Hopefully, everything from  $P + 1$  to  $P + n$

## Reading the sequence

- DNA from the sequencing reaction is purified via ethanol precipitation
- DNA is resuspended in deionized formamide
- Plate is loaded into the automated sequencer

## Automated sequencing

- Capillary array contains polyacrylamide gel

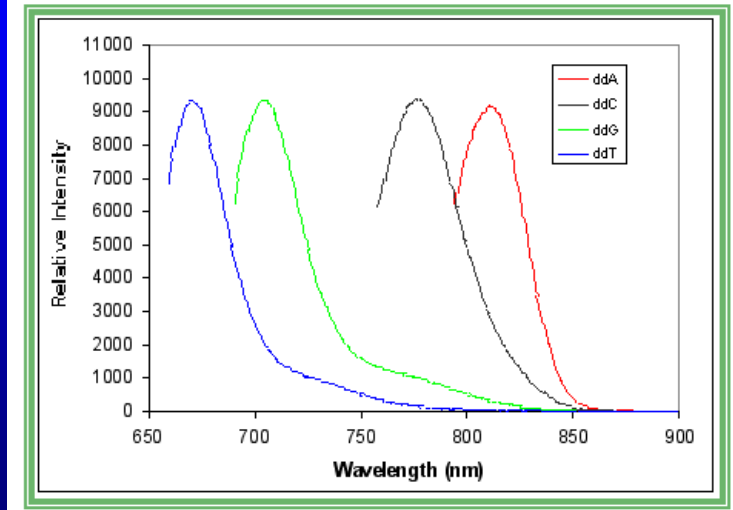


- DNA fragments migrate through gel by electrophoresis
- Separate by size

## Automated sequencing

- Capillary passes through a laser
- Each dye fluoresces a different wavelength when excited by the laser
- Fluorescence is detected by a CCD

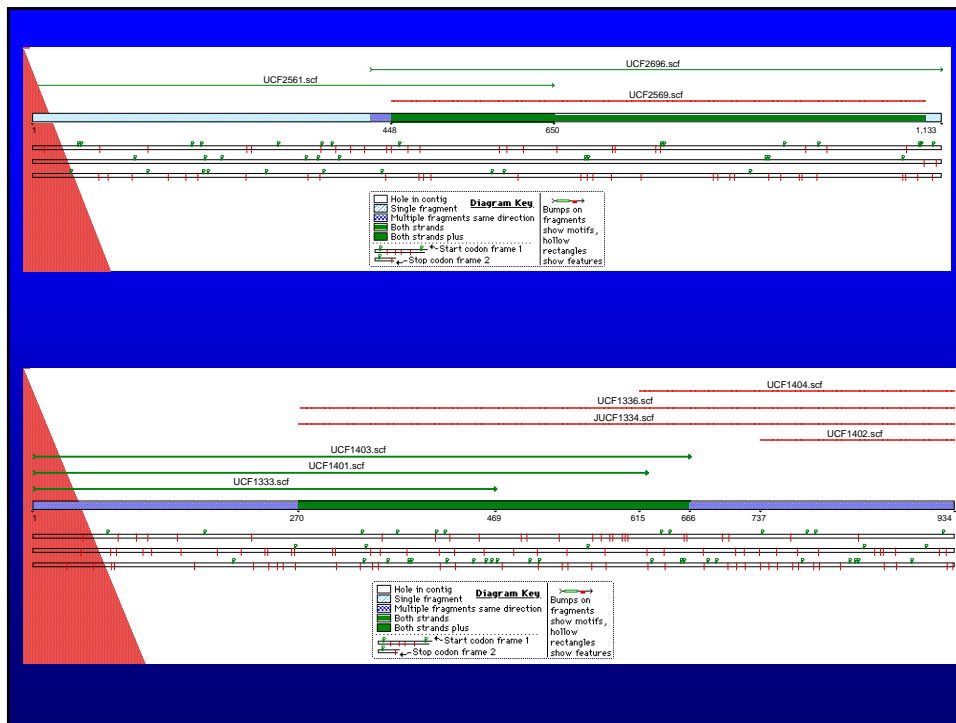
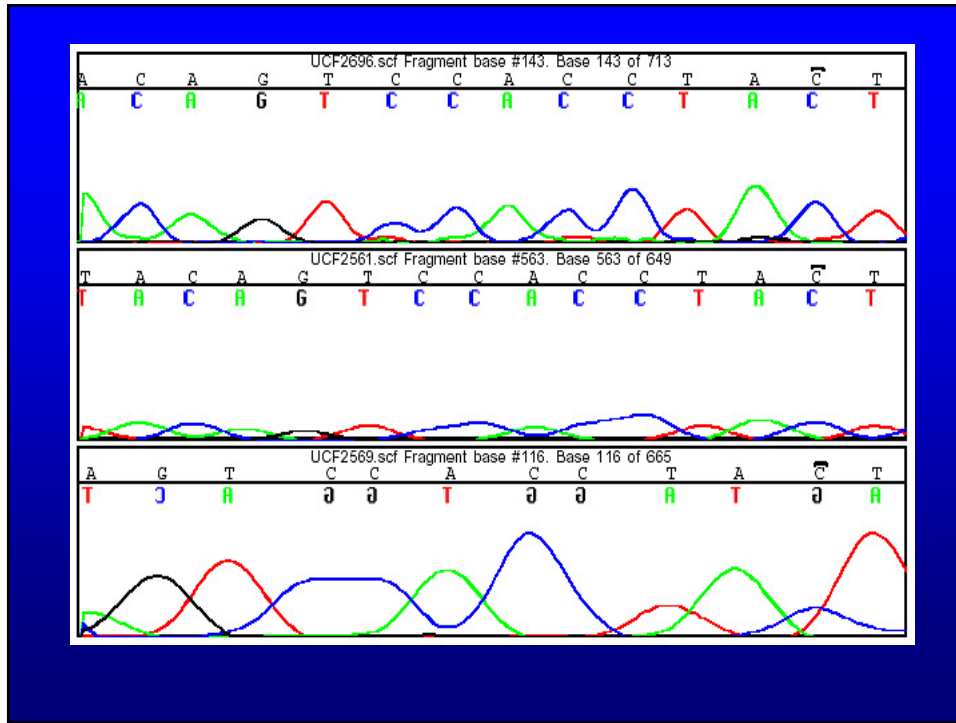
## Fluor Emission Spectra



## Automated sequencing

- Fluorescences are processed into an electropherogram
- Base “calls” made by sequencing software, but can be analyzed manually





## **NCBI – National Center for Biotechnology Information**

- <http://www.ncbi.nlm.nih.gov/>
- Literature databases
- Entrez databases
- Nucleotide databases
- Genome resources
- Analytical tools

## **Literature databases**

- PubMed – searchable citation database of life science literature
- PubMed Central – digital versions of life science journals
- Bookshelf – online versions of textbooks
- OMIM – catalog of human genes and genetic disorders
- PROW – Protein Reviews On the Web – reviews of proteins and protein families

## Entrez databases

- **System for searching several linked databases:**
  - PubMed
  - Protein sequence databases
  - Nucleotide sequence databases
  - Genome databases
  - Pop sets
  - Books

## Nucleotide databases

- **GenBank - annotated collection of all publicly available nucleotide and amino acid sequences**
- **SNPs - Single base Nucleotide Polymorphisms - substitutions and short deletion and insertion polymorphisms**
- **ESTs - Expressed Sequence Tags - short, single-pass sequence reads from mRNA**



## Genome resources

- Whole genomes of over 800 organisms
- Others in progress
- Viroids, viruses
- Plasmids
- Bacteria
- Eukaryotic organelles

## Genome resources

- Eukaryotes
  - Yeast
  - Fruit fly
  - Zebrafish
  - Human
  - *C. elegans*
  - *Rattus, Mus*
  - *Plasmodium*
  - Plants

## Analytical tools

- Sequence analysis tools
- Macromolecular and 3-dimensional structure analysis
- Software downloads
- Citation searching
- Taxonomy searching
- Sequence similarity searching – BLAST

## Where are we now??

- Kelly has shown you PCR....
- Matt has explained sequencing...
- Now we must use BLAST with our sequence to determine if we have the correct:
  - Gene
  - Animal

# BLAST

- **Basic Local Alignment Search Tool**
- **Similarity Program**
  - Compares input sequences with all sequences (protein or DNA) in database
  - Each comparison given a score
    - Degree of similarity between query (input sequence) and sequence that it is being compared to
    - Higher the score, the greater the degree of similarity

## BLAST, cont'd

- **Significance of each alignment composed as an E-value**
  - The number of different alignments with scores equal to or greater than the given score that are expected to occur in a database search by chance
  - The lower the E-value, the more significant the score